

# Systemic Small Molecule TREX1 Inhibitors to Selectively Activate STING in the TME of Metastatic Disease

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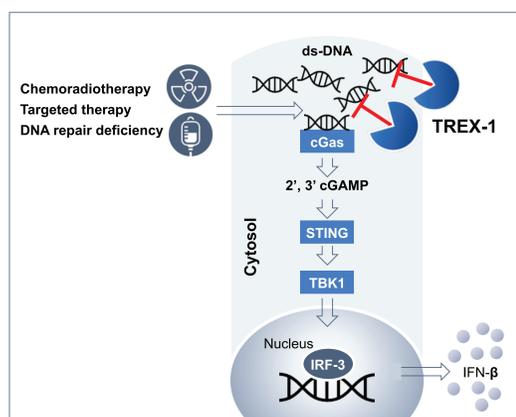


## ABSTRACT

Genetic evidence from human disease and mouse genetic knock-out studies identify the Stimulator of Interferon Genes (STING) pathway as a critical innate immune sensor for the development of immunity. Tumor cells can evolve to avoid immune recognition through inactivating the STING pathway by diverse mechanisms, indicating that it is important to generating tumor-specific immunity. However, the clinical activity of STING agonists given by intratumoral (IT) administration has not compared well to preclinical studies. The scientific hypothesis supporting these first clinical studies is that localized CD8<sup>+</sup> T cell priming would have activity against distal non-injected tumors, but findings that tumors in advanced malignancies have unique antigenic repertoires suggests that this approach may have limited activity against distal tumors. Global innate activation in metastases may therefore be necessary to prime a broadly active CD8<sup>+</sup> T cell population targeting diverse antigens, in addition having the benefit of reversing the immune suppressive tumor microenvironment (TME). However, ubiquitous expression of STING presents a significant challenge to achieving a therapeutic index with systemic delivery of direct STING agonists. Selective activation of the STING pathway may be achieved through targeted inhibition of TREX1, a cytosolic DNA exonuclease that modulates cGAS/STING signaling. Expression of TREX1, in contrast to STING, is increased in tumor cells due to elevated levels of cytosolic DNA resulting from genetic instability, DNA repair mutation, inflammatory mediators or DNA-modifying anti-cancer therapies. These observations provide the principal scientific rationale to selectively activate the STING pathway in metastatic disease through targeted inhibition of TREX1. Utilizing published TREX1 X-ray crystal structures to guide medicinal chemistry, we discovered small molecule inhibitors of TREX1 and transformed these molecules from >100 μM leads into a series with drug-like physicochemical properties and picomolar potency against both human and mouse TREX1. We evaluated the activity of lead molecules in cell-based assays, in which TREX1 inhibition enhanced cGAS/STING signaling, and advanced molecules with desired pharmacokinetic profiles to mouse tumor models. We observed significant anti-tumor activity in mice with CT26 tumors given a combined therapy of low dose doxorubicin to induce dsDNA breaks and increase TME TREX1 expression along with lead series TREX1 inhibitors. Recognizing that TREX1 is a DNA repair enzyme, we also showed that TREX1 inhibitors were cytotoxic in DNA repair deficient human tumor cell lines, informing advancement of this new class of STING therapeutics as a clinical approach to both activate the cGAS/STING pathway to initiate immune recognition, as well as to inhibit DNA repair orthogonal to existing tumor-driver DNA repair mutations.

## INTRODUCTION

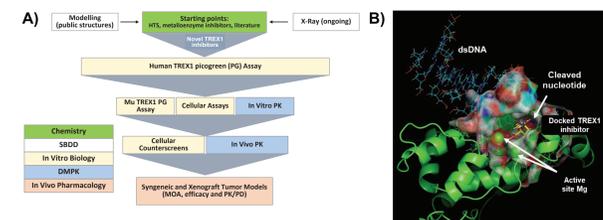
### Drugging the STING Pathway Through Targeting TREX1



- STING is a critical innate immune sensor for the development of immunity
  - STING activating mutations result in profound immune inflammatory diseases
  - Malignant cells evolve to inactivate STING by diverse mechanisms
- TREX1 is a negative regulator of STING
  - Cytosolic dsDNA 3'-5' non-processive exonuclease
  - DNA base and nucleotide excision repair enzyme
  - Reduces cGAS-STING activation by DNA
- TREX1 expression can be upregulated in the TME
  - Tumor genetic instability and DNA damaging therapies increase cytosolic dsDNA and induce TREX1
  - TREX1 upregulation protects tumor from immune recognition
- TREX1 inhibition activates the STING pathway
  - Enhanced TREX1 expression in tumors vs healthy tissues should improve therapeutic index compared to direct STING agonists

## APPROACH

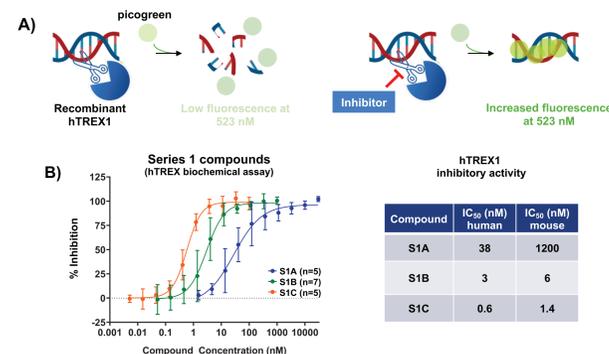
Figure 1. Development of Selective TREX1 Inhibitors



Approach to develop potent and selective TREX1 inhibitors. A) Screening funnel schematic illustrating steps taken to select TREX1-specific inhibitors. Steps shown are iterative based on experimental results; B) Lead Series molecules are predicted to bind in the TREX1 active site. Our docking model (based on a published mouse TREX1/DNA co-crystal structure) aligns with ongoing SAR (not shown).

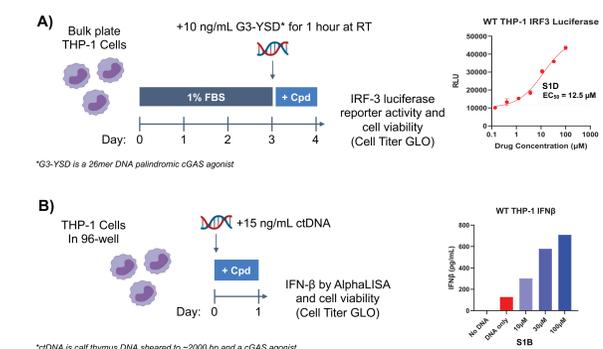
## RESULTS

Figure 2. Activity of a Lead Series TREX1 inhibitors in biochemical assay



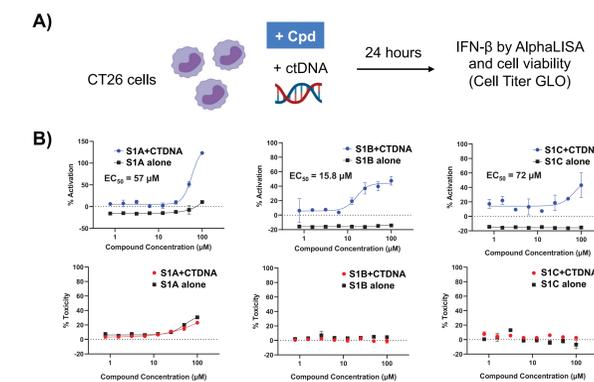
Potent inhibition of TREX1 inhibitor compounds in biochemical fluorescent assay A) Assay schematic illustrating combination of compound with dsDNA template labeled with picogreen to assess compound TREX1 inhibitory activity. Enzymatic assays were conducted with 8 ng/mL TREX1 protein in buffer with DDT incubated at 25 °C for 30 min with compound at desired concentrations followed by addition of dsDNA template at 50 nM final concentration, mixing, and incubation at 25 °C for 5 min. Inhibition of DNA degradation was measured by fluorescence at 520 nm upon 480 nm excitation immediately following the addition of EDTA stop solution containing picogreen. B) Human TREX1 inhibitory activity of lead molecules determined over 10-point dose range. Table shows IC<sub>50</sub> values of lead molecules against both human and mouse TREX1.

Figure 3. Activity of a Lead Series TREX1 inhibitors in THP-1 reporter cell lines



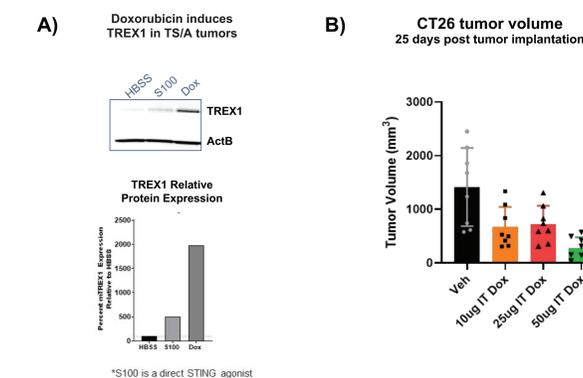
A) TREX1 inhibitors increase STING activation in THP-1 reporter cell lines. Schematic showing process for assessment of inhibitor activity through measurement of IRF-3 activation of luciferase reporter. THP-1 cells were batch transfected with 10 ng/mL G3-YSD in Lyovect for 1 h at RT. Cells were added to 96-well plates and incubated for 24 h at 37 °C following addition of drug. Reporter expression was measured in cell supernatants with Quanti-Luc. Example of EC<sub>50</sub> results with TREX1 inhibitor determined over 6 different dose levels. B) TREX1 inhibitors increase production shown of IFN-β in THP-1 reporter cell lines. 96-well plates were prepared as in (A) except DNA stimulation was with 15 ng/mL sheared ctDNA in Lyovect. IFN-β expression was measured in cell supernatants by AlphaLISA. Example shown of IFN-β production in the presence of TREX1 inhibitor at different concentrations.

Figure 4. Activity of a Lead Series TREX1 inhibitors in CT26 tumor cells



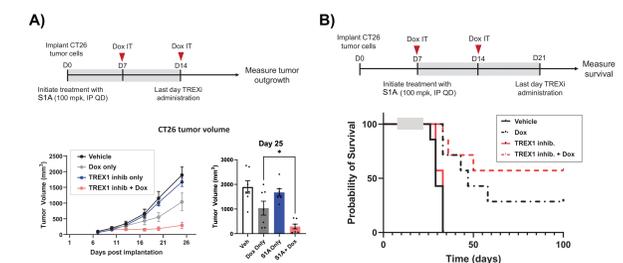
A) TREX1 inhibitors increase IFN-β production in DNA-stimulated CT26 tumor cells. Schematic showing process for assessment of inhibitor activity through measurement of IFN-β production. B) Cells were prepared in a 96-well format and stimulated with 50 ng/mL ctDNA in Lyovect and drug in DMSO for 1 h as in Fig 3. IFN-β expression was measured in cell supernatants by AlphaLISA. Cell viability was determined using Cell Titer GLO.

Figure 5. Chemotherapy-mediated induction of TREX1 in tumors



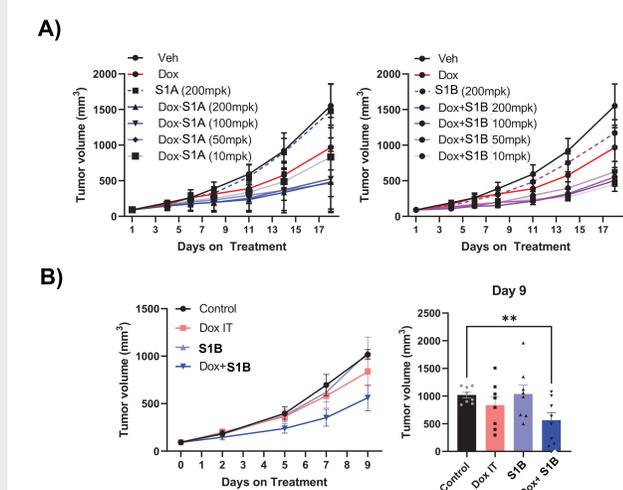
Subtherapeutic doses of Doxorubicin induces dsDNA breaks and activates TREX1 expression. A) TSA tumors were injected with Vehicle, 25 μg Doxorubicin (dox) or with S100 (direct STING agonist) and 48 h later were prepared for Western blot and qPCR analyses by dissociating tumor in collagenase and benzonase at 37 °C. Processed tumors were filtered and aliquoted. Lysates were prepared for Western blot in RIPA buffer with protease/phosphatase inhibitor cocktail (PIC). After filtering and aliquoting tumors, lysates were prepared for western blot by resuspending the tumor aliquot for western blot in 500 μL RIPA buffer with PIC. 50 μg of protein was electrophoresed on 4-12% bis-Tris gels. TREX1 and Actin-B were detected with specific mAbs (Cell Signaling Technology), diluted 1:1000 and 1:2000, respectively. Gel images were obtained with the Azure system. RNAeasy (Qiagen) was used to prepare RNA for qPCR. B) Groups of 8 Balb/c mice bearing flank CT26 tumors were given two intratumoral (IT) injections of dox separated by 1 week at the doses shown in the Figure, once tumors were ca. 100 mm<sup>2</sup> at 11 days post cell implantation.

Figure 6. Anti-tumor activity of TREX1 inhibitors in CT26 tumor model



TREX1 Lead Series molecule therapy results in significant anti-tumor efficacy in combination with subtherapeutic doses of doxorubicin. A) Groups of 7 Balb/c mice were implanted with CT26 tumor cells and on D0 were given 100 MPK of S1A TREX1 inhibitor by IP injection, QD for 14 days. Tumors on D7 and D14 were given 10 μg of dox intratumorally. Tumor growth was measured every 3 days until D25. B) Groups of 7 CT26 tumor-bearing Balb/c mice were treated with vehicle, Dox on days 7 and 14, or 14 days of S1A at 100 MPK administered IP QD beginning on D7 when tumors were ca. 100 mm<sup>2</sup> and monitored for survival. Significance (combination versus dox alone) was determined by Student's 2-sided t test.

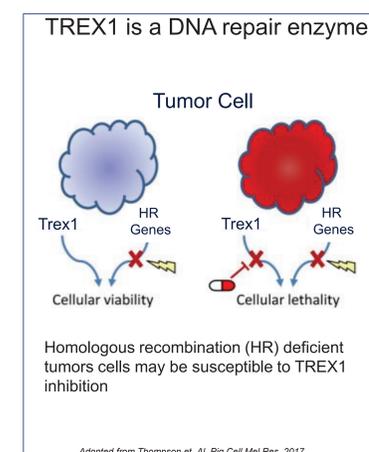
Figure 7. Dose response of TREX1 inhibitors and activity in MC38 tumor model



A) Anti-tumor efficacy correlates with TREX1 inhibitor IC<sub>50</sub> value. Groups of 8 CT26 tumor-bearing Balb/c mice were treated with Vehicle, Dox on Days 7 and 14, or a subtherapeutic dose of Dox in combination with TREX1 inhibitors S1A or S1B given IP over a dose range between 10 and 200 MPK for 18 days. Tumor growth was measured every 3 days during TREX1 inhibitor therapy. Relative in vivo efficacy correlates with in vitro activity (Figure 2). B) TREX1 inhibitors have comparable anti-tumor activity in the MC38 tumor model. Groups of 8 C57Bl/6 mice bearing ca. 100 mm<sup>2</sup> tumors were treated with vehicle, once with Dox, or once with Dox and 200 MPK of S1B given IP for 9 days and tumor growth was monitored. Significance was determined by Student's 2-sided t test. (\*\*: p<0.01)

## SUMMARY & NEXT STEPS

- We have developed several series of potent TREX1 specific inhibitors
  - Docking model based on published mTREX1/DNA X-ray crystal structures strongly aligns with SAR
  - pM inhibition of human TREX1 in biochemical assays
  - Enhanced activation of STING pathway in DNA-stimulated human and mouse cells
  - Significant therapeutic anti-tumor efficacy and survival when combined with the DNA damaging agent doxorubicin
- Two features of TREX1 biology can be targeted clinically
  - Activation of cGAS/STING pathway selectively in the TME
  - Synthetic lethality partner in DNA repair mutant tumors (preliminary results not shown)
- Unique approach to selectively activate the STING pathway in the TME with a systemically administered TREX1 inhibitor
- Ongoing SAR activities with lead series compounds together with immunologic and tumor lethality MOA studies towards selection of a Development Candidate for IND-enabling activities are ongoing



Adapted from Thompson et al. *Al. Pig Cell Mel Res.* 2017